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# Bicyclo[3.2.1]-DNA: A DNA Analog Containing a Rigid Backbone and a Flexible Base-Pairing Region

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## BICYCLO[3.2.1]-DNA: A DNA ANALOG CONTAINING A RIGID BACKBONE AND A FLEXIBLE BASE-PAIRING REGION.

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**ABSTRACT**: Bicyclo-[3.2.1]-pyrimidine nucleoside phosphoramidites were synthesized from a common bicyclo-[3.2.1] "sugar" intermediate. Oligomers up to 20 nucleotides in length were synthesized successfully. UV melting curve and thermodynamic analysis of these oligomers reveal stable, antiparallel duplexes

We designed the nucleoside analog bicyclo-[3.2.1] system to lock the  $\gamma$  and  $\delta$  torsion angles to that observed in a B-DNA helix while allowing torsion angles  $v^0$  and  $v^4$  to be conformationally flexible, to which a base may be attached. With the goal in mind of incorporating this analog into nucleic acid oligomers, the appropriate pyrimidine phosphoramidite monomers 1 and 2 were synthesized. \(^1

Rigid 
$$\delta$$

Base

O

DMTO

Base

1 Thymine

2 Cytosine

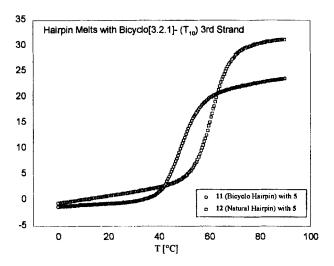
FIG. 1

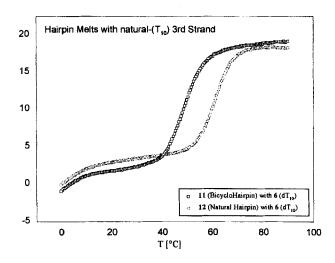
Fully modified oligonucleotides up to 20 bases in length were successfully synthesized using standard automated solid-phase methodology.

3	5'-T <sub>5</sub> - <u>T</u> -T <sub>4</sub> -3'	9	5'-TCT <sub>2</sub> C <sub>3</sub> TCTC	CT5CT2	Γ-3'
4	5'-T <sub>3</sub> - <u>T</u> <sub>4</sub> -T <sub>3</sub> -3'	10	5'-TCT <sub>2</sub> C <sub>3</sub> TCTC	$\Gamma_5CT_3-3$	
5	5'- <b>T</b> <sub>9</sub> T-3'	11	$5'-A_{10}C_3T_9T-3'$		
6	5'-T <sub>10</sub> -3'	12	$5'-A_{10}C_3T_{10}-3'$	15	5'-A <sub>3</sub> GA <sub>5</sub> GAGAG <sub>3</sub> AAGA-3'
7	5'- <u>T</u> 5-( <u>CT</u> ) 4 <u>C</u> T -3'	13	5'-A <sub>10</sub> -3'	16	5'-AGAAG3AGAGA5GA3-3'
8	5'-T <sub>5</sub> -(CT) <sub>5</sub> -3'	14	5'-(AG) 5A5-3'		
•	= Ricyclo-13 2 11-cytosine T =	Ricyclo-	[3.2.1]_thumine all other	rc are natur	ral nucleotides

Entry	Duplex	T <sub>m</sub> (° C)	$\Delta T_{\rm m}$ / mod.	ΔH (kcal/mol <sup>-1</sup> )	ΔS (cal K <sup>-1</sup> mol <sup>-1</sup> )	ΔG <sup>25° C</sup> (kcal/mol <sup>-1</sup> )
A	3•13	25.5	-7.5	nd	nd	nd
В	4•13	13.3	-4.9	nd	nd	nd
С	5•13	5.0	-3.1	nd	nd	nd
D	6•13	33.0	0	nd	nd	nd
E	7 <u>•14</u>	37.6	-1.3	$-54.2 \pm 0.4$	-149.4 ± 1.3	$-9.7 \pm 0.8$
F	8-14	54.9	0	-101.1 ± 0.4	-281.6 ± 1.3	-17.1 ± 0.7
G	9•15	48.1	-0.7	-94.5 ± 0.8	$-268.0 \pm 2.4$	-14.7 ± 1.6
Н	10•15	61.9	0	-150.7 ± 1.2	$-420.9 \pm 3.5$	-25.2 ± 1.8
I	9•16	~15	n/a	n/a	n/a	n/a
J	10•16	24.6	n/a	n/a	n/a	n/a
K	11	42.6	-1.4	-47.6 ±0.3	$-150.2 \pm 0.9$	$-3.3 \pm 0.6$
L	12	56.9	0	-61.7 ± 0.1	-187.2 ± 0.4	$-6.4 \pm 0.3$

All UV melts were obtained in 10 mM Na-Cacodylate, pH 7.0, 1 M NaCl. Oligonucleotide concentration: A-D 4  $\mu$ M, E-L 2 $\mu$ M.





Synthesized oligonucleotides were analyzed by UV melting curves to determine the effects of bicyclo-[3.2.1]-nucleosides on duplex stability and binding orientation, and thermodynamics were determined from curve-fitting analysis.

To assess the propensity of bicyclo-[3.2.1]-DNA to bind to either natural or hybrid duplexes in the pyrimidine triplex motif, modifed decamer 5 and natural decamer 6 were targetted to hairpins 11 and 12. In each case no denaturation of third strand is visible. However, natural decamer 14 was able to bind both the natural and hybrid hairpins, demonstrating that while bicyclo-[3.2.1]-DNA is unable to bind as a third strand, its presence in a duplex does not preclude triplex formation by a natural third strand.

#### REFERENCE

1 for synthetic conditions see: Egger, A., et al., Helv. Chim. Acta, 1998, 81, 734. Epple, C., Leumann, C. Chem. Biol., 1998, 5, 209.